

Elegans Newsletter

Notes from the „elegans study-group“ of the DKG

- 3/2001 -

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Dear *elegans*-friends,

I am happy to welcome a new member in the group – Christian Ropkopf (DKG 40), who is a well known *Cynolebias* specialist. Currently, Christian is maintaining the following *elegans*-populations:

A. christyi HZ 85/8

A. lamberti "BSWG 97/9"

A. rectogoense PEG 95/16

On the other hand, I had to remove Markus Thun, Horst Tomczyk, Daniele Cerri und Jan Wester from the list, because they did not ever respond to one of my letters.

Dirk Ude's email address changed (see cover)

1. 2001 DKG convention in Augsburg

The meeting of the *elegans* study-group in Augsburg was well attended. Especially surprising was the appearance of three new interested people, which were:

Wilfried Pütz, DKG 33 (has COG, CON and ROG)

Hartmut Reusch, DKG 165 (interested in *A. schioetzi*)

Hans-Gerd Tschuch, DKG 396 (distributed *A. cognatum* „Kinsuka“)

Furthermore, the following group members were present:

Jürgen Heidler

Hans Kämpf

Axel Schwekendiek

Dirk Ude

Rüdiger Wagner

It was a real pleasure to see the faces to the names and email addresses finally. I really hope there will be more of you coming next year.

The fishes from Tschuch were of superior quality and Dirk Ude and myself were able to obtain lots of fry in the meantime.

Now the most important news from Augsburg:

I) The ENL shall be published just twice a year (after the convention which is in may each year and around christmas)

II) the species conservation program shall be pursued as planned. In chapter 2.) the species coordinators are named. However, there has still NO priority list being issued. Currently, the decision about which populations have to be given priority in preservations must be made by the species coordinators. Maybe we can develop a comprehensive list next year?

III) next year we will send the entire list of elegans-species maintained by us to Jan Poetke for integration into the DKG’s annual species maintenance list and not seperately by each person individually.

2. Updates Species Maintenance List

Unfortunately, not everybody has informed me about updates of his fishlist since my last request from may. So please check the list and give me any corrections or updates soon.

Ident-No.	Species – Population	Name
427	A. chauchei „Masèpe“	Haggerty Kämpf Kraska Schwekendiek Wagner
4249	A. chauchei „Obeya“, RPC 91/6	Haggerty Hölzer
430	A. chauchei „Oltombo“, RPC Z 96/5	Haggerty Hölzer Kraska Ude Wagner
432	A. christyi „Aquariestamm“ A. christy “alter DDR Stamm”	Hölzer Heidler
437	A. christyi „HZ 85/8“	Haggerty Harder Hölzer Kämpf Kraska Lehmann Ott Roßkopf Türk

3993	A. christyi “HZ 85/20”	Wagner
436	A. christyi “HZ 85/22”	Kämpf Kraska
442	A. christy “Lobaye”	Conrad Kämpf Schwekendiek
432	A. christyi “Aquariestamm”	Hölzer
499	A. cognatum “Z 82/2”	Haggerty Heidler Kraska McNealy
500	A. cognatum “Gilima”, Z 91/3	Haggerty Ott
484	A. cognatum „Kenge“, CI Z 93	Haggerty Hölzer Kämpf Kraska
489	A. cognatum “Kinshasa”	Weber
1441	A. spec. aff. cognatum “Lake Fwa”	Haggerty Hölzer Kämpf Lehmann McNealy Schwekendiek
490	A. cognatum “Kinsuka”	Harder Schwekendiek Ude Wagner
499	A. cognatum “Z 82/2”	Kraska Wagner
4251	A. cognatum “CI 2001”	Türk
501	A. congium	Lehmann McNealy
505	A. congium „Z 82/17“	Haggerty Heidler Kämpf Kraska Lehmann Ott Tomcyk Weber
516	A. decorsei „Bangui“, KCF 5	Kämpf Kraska Ude
442	A. spec. aff decorsei “Lobaye”	Kraska
518	A. decorsei “Kapou I”, RCA 91/1	Haggerty Hölzer Kämpf Kraska Lehmann McNealy

		Pinto
517	A. decorsei “RCA 82/3”	Haggerty Hölzer Kraska Schwekendiek
	A. decorsei “RCA 91/3” (?)	Addis
	A. lamberti “BBA/GAB 90/1”	Addis Brown
848	A. lamberti „BSWG 97/9“	Lehmann McNealy Pelletier Roßkopf Türk
854	A. lamberti „G 88/7“	Haggerty Kämpf
855	A. lamberti „G 92/4“	Kraska
860	A. lamberti “GEB 94/7”	Hölzer Kämpf
4250	A. lamberti „GHH 97/11“	Weber
873	A. lamberti “PEG 96/4”	Kämpf
878	A. lamberti “TDK 97/17”	Addis Lehmann Weber
881	A. lefiniense “CI 96”	Conrad Haggerty Hölzer Kraska Ott Ude
986	A. margaretae “HZ 85/14”	Conrad Haggerty Kämpf Kraska Ott
1392	A. spec. “oyo” ,RPC 91/8	Addis Drake Haggerty Hölzer Kämpf Kraska Lehmann Ott Schwekendiek Ude Wagner
1192	A. polli	Hölzer
1195	A. polli “N’jili”, CI Z 93 (Type loc.)	Wagner
1194	A. polli “CI 95”	Kämpf Schwekendiek
4253	A. polli “CI 99”	Haggerty Ude

4252	A. polli “DDR-Stamm”	Wagner
1272	A. rectogoense „BBA/GAB 90/27“	Addis Brown Conrad Kämpf Lehmann Weber
1270	A. rectogoense “G 92/5”	Haggerty Kämpf
1267	A. recotgoense „G 92/11“	Haggerty Kraska Tomcyk Wagner
1275	A. rectogoense „GHP 80/30“	Conrad Haggerty Harder Hölzer Lehmann
1276	A. rectogoense “PEG 95/16”	Hölzer Kämpf Lehmann Ott Roßkopf Schwekendiek
1319	A. schioetzi „?“	Drake Hölzer McNealy Pelletier
1321	A. schioetzi „C 85/1“	Haggerty Ude
1336	A. schioetzi “Z 91/1”	Hölzer Kämpf Ude Weber
1389	A. spec. “Epoma” “RPC 91/1”	Haggerty Heidler Hölzer Kraska Ott Wagner
1381	A. spec. “Madimba”	Schwekendiek

Further to the list, Jan-Luc Claisse (KCF) gave me his fish list:

- No. 437 *A. christyi* HZ 85/8
- No. 442 *A. christyi* Lobaye
- No. 505 *A. congicum* Z82/17
- No. 986 *A. margaretae* HZ 85/1
- No. 1286 *A. rectogoense* PEG 95/16

Also Alexey Rak sent me his list (all fishes are from Dennis Haggerty):

- No. 1321 *A. schioetzi* C85/1
- No. 505 *A. congium* Z82/17
- No. 517 *A. decorsei* RCA82/3
- No. 1392 *A. sp.oyo* PRC91/8

Wolfgang Vespermann (DKG 167) called me in february and that time had *A. lefiniense* CI 96, *A. chauchei* „Oltombo - RPC Z 96/5“ and *A. christy* (rot) „HZ 85/22“. I had no contact to him since.

3. Species Coordinators

This is the current list of species coordinators for our species maintenance program:

Harald Hölzer for *A. christy*

Jürgen Heidler for *A. cognatum* and *A. congium*

Monty Lehman for *A. rectogoense*

Rüdiger Wagner for *A. chauchei*, *A. polli* and *A. spec.* Epoma

Ewald Türk for *A. lamberti*

Dirk Ude for *A. schioetzi*, *A. lefiniense* and *A. decorsei*

Axel Schwekendiek for *A. spec.* Lake Fwa, *A. spec.* Madimba and *A. spec.* Oyo

Therefore we still have nobody *A. margaretae*, but I would suggest to integrate it to *A. christyi*. Then we would be set for the moment.

It would be great if the species coordinators would contact the newly interested people Vespermann, Tschuch, Pütz, Claisse (jlclaisse@met.wallonie.be) and Rak (alex.rak@mpi-dortmund.mpg.de) about their species. This would mean Dirk Ude, Rüdiger Wagner, Harald Hölzer, Jürgen Heidler and myself. However, also the breeders are encouraged to contact the respective coordinators.

I got some questions about the tasks of the species coordinators recently. This seems to be somewhat unclear. Even if I don't want to define any tasks (I think this should be the responsibility of the coordinators themselves – what do they think is important?), I compiled some of my suggestions in the following list:

- maintain contact to all the breeders from the list of the respective species/population
- decision about the population(s) which should be put at highest priority in species preservation
- maintain species lists
- development of vital populationis (share and trade individuals)
- take care for substitutes in case someone drops out of the program – especially keep the populations in the hobby and find someone who is willing to take over critical strains
- write updates (problems, successes) for the ENL every 6 months (involve other members)
- write reviews or fact sheets for the species/population(s) in question
-your suggestions here next time ???

Especially, I would like to encourage everybody to get in touch with the respective coordinator for your own species. Don't wait until you are contacted. It must not always be the coordinator who makes the first step. Just stay in touch from time to time and get involved regularly.

4. Hatching assistance

Rüdiger Wagner suggested to collect a list of methods to induce hatching. The following reports reached me:

Christian Roßkopf:

My method to get the fish out of the eggs is the following:

I keep all eggs on peat moss. As soon as the eggs developed, I pour cool water on them. 16-20°C depending on species and also depending on breeding temperature. What does not want to hatch stays inside. This is a somewhat provocative method. However, In my opinion the “bad” hatching may be genetically determined. If you always force the eggs to hatch, you will probably raise “bad” hatching strains over time. With my method I observed increased hatching of the next generation compared to the parents. Of course, this

methods only work if you get enough eggs and if the gender ratio is somewhat equal. If you were dependent on every single hatched fry because you simply do not get enough eggs or if you have to pray for every fish: “hope this is the one male to my 100 females” then you have a problem and have to do something. In this case you should ask yourself if there is a problem with egg storage which might be the cause of bad hatching rates (look at the problem with *S. flauvicaudatus* form www.cynolebias.de). I always had bad hatching rates with “*Roloffia*” schmitti until I stopped collecting eggs and simply raised the fry in the parents tank. After 2 months I raised more than 100 fry this way. Maybe the development of the fish could be disturbed by collecting eggs or the storage of the eggs influences the hatching behaviour in some way?

Thomas Kraska

There was an article in the last DKG journal about hatching assistance. I follow this. In the first instance I try cool or cold tap water. Sometimes I use oxygen pills. Especially with *Dolichopterus* I hardly get any belly sliders this way. In difficult cases I use dry food. I just put it on the water surface. Just a little so the water does not get too bad too fast. This builds infusoria quickly and the oxygen drops. So I get difficult strains to hatch. I was a little uncertain about this method – but it works. If nothing helps you should look at the breeding pair. Maybe they don't match or are from incompatible strains?

Axel Schwekendiek

In first instance, I always collect the eggs from mop just as I find some. Either I separate male and female for a week or I just go ahead and keep the mop in the species tank, looking for eggs occasionally. At first, I keep the eggs exactly three weeks on wet peat moss at 20-23°C. For hatching, I put them in 18°C cold water and wait two days. If nothing happened, I put the eggs back to moss for an additional week. Again 18°C cold water and two days. If nothing happens, dry food on the water surface as Thomas Kraska did. Wait again two days, which in most cases helps. If not, try again whole procedure but keep for two weeks only on peat. This worked in the past for all species (for *epiplatys* and *Fundulopanchax* too).

It would be nice if the species coordinators could compile a list of eggs-storage times (incl. storage temperatures) for their populations for the next ENL.

Thøger Harder

My method for breeding is very simple. I keep the parents in a 60 l aquarium with rainwater, a lot of *Javamos* (*Vesicularia dubyana*) and some peat on the bottom. I remove the parents, when I see young or let the fry grow up together with the parents.

5. Misc

A. christyi HZ 85/20 back to Germany

Rüdiger Wagner was able to bring back *A. christyi* HZ 85/20 to Germany.

A. christyi Lobaye and *A. decorsei* RCA 82/3 identical ?

Thomas Kraska believes, *A. christyi* „Lobaye“ and *A. decorsei* „RCA 82/3“ might be identical because of color patterns. I too have both strains in my basement and must admit that they look pretty close. However, I would warn to judge two populations the same species just because of color patterns. This is not enough for separating species or putting them together. We should discuss this topic in more detail and/or undertake some cross-breeding experiments first. If the results support Kraska's view, we would have to redefine *A. christyi*, *A. decorsei* or both. Is anyone aware of first descriptions for *A. christyi* and/or *A. decorsei* ???

A. chauchei Marsepe dying

Recently, I encountered serious problems with my *A. chauchei*. After I could raise several males (not a single female, though), I received some females from friends and put them all together into a 15 gallon tank. B.T.W. Rüdiger Wagner mostly raised females ☺ Now, the males all died but no female. First, the largest males died (3-5 a day) and later on the younger ones. Every day the same – 3-5 fish dead. This continued even after 100% water change and after a week I had lost all males and still have the females. I could not detect any disease or injuries on the males and have absolutely no clue what happened. Any suggestions would be very much appreciated, since I still have lots of fry in the pipeline (in a different tank and still not colored).

Problems mit *A. polli* CI 95

Also with *A. polli* I have serious problems. In Augsburg I received another pair of the population CI 95 from Hans Kämpf, additional to my lone male. The new pair immediately produced 14 eggs of which not a single one hatched.

Unfortunately, only 3 fry hatched. The “new” males died soon and with the “old” male I just get fungused eggs (only one out of 300 did not fungus). Could this be due to incompatible individuals or a semi-sterile male? Did anyone make similar observations, maybe with other strains?

Looking for *A. lamberti*

Ewald Türk is looking for *A. lamberti* – only with location code - except for BSWG 97/9

contact:

etuerk@tuerk-tech-mall.net

Tel. 06103 9604 12

only a.m. 10-11 Uhr, othwise answering machine

Fax 06103 9604 15

That’s it for this time. I hope to get your feedback on the questions raised in this issue.

Best wishes and good luck with the breeding,

Axel

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